EUROPEAN PATENT APPLICATION

(43) Date of publication: 26.08.1998 Bulletin 1998/35

(51) Int Cl.6: **G01N 33/50**, G01N 33/569

- (21) Application number: 98301123.0
- (22) Date of filing: 16.02.1998
- (84) Designated Contracting States:

 AT BE CH DE DK ES FI FR GB GR IE IT LI LU MC

 NL PT SE

 Designated Extension States:

 AL LT LV MK RO SI
- (30) Priority: 21.02.1997 US 38914 P
- (71) Applicant: SMITHKLINE BEECHAM
 CORPORATION
 Philadelphia Pennsylvania 19103 (US)

- (72) Inventor: Del Vecchio, Alfred King of Prussia, PA 19406 (US)
- (74) Representative: Crump, Julian Richard John fJ Cleveland, 40-43 Chancery Lane London WC2A 1JQ (GB)
- (54) Use of HSV-1 UL-15 and VP5 in identifying anti-viral agents
- (57) The present invention provides a method for identifying compounds having antiviral activity against members of the herpes virus family. This method involves the identification of compounds which inhibit the

interaction between the gene products of the herpes simplex virus-1 UL15 gene and the UL19 gene (VP5). Also provided are compounds identified by the method of the invention.



Description

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Field of the Invention

This invention relates generally to the field of anti-viral agents, and particularly to methods of identifying anti-viral agents active against members of the herpes family.

Background of the Invention

Members of the Herpesviridae family include the human pathogens herpes simplex virus types 1 and 2 (HSV-1 and HSV-2), varicella-zoster virus (VZV), human cytomegalovirus (HCMV), human herpes viruses 6 and 7 (HHV-6 and HHV-7), Epstein-Barr virus (EBV) [reviewed in The Human Herpesviruses, ed. B. Roizman et al, Raven Press, NY (1993), pp. 1-9], and the Kaposi-sarcoma herpesvirus (KSHV) or HHV-8 [P. S. Moore et al, J. Virol., 70: 549-558 (1996)]. These viruses all have a large (approximately 100-270 kilobase pairs), linear, double-stranded DNA genome [reviewed in D. J. McGeoch, Annu. Rev. Microbiol., 43:235-265 (1989)]. During productive infection, viral DNA replication occurs via a proposed rolling circle mechanism producing concatameric copies of the viral genome. During maturation of the virion particle, a capsid particle self assembles, and the newly replicated viral DNA is cleaved into a unit length genome and subsequently packaged into the empty capsid (B-capsid) [See, The Human Herpesviruses, cited above, at pp. 11-68]. The processes of DNA cleavage and packaging of the viral DNA into the capsid are believed to be tightly coupled, as no viral mutants have yet been identified which cleave, but do not package DNA. Furthermore, this process of DNA cleavage and packaging is dependent upon the formation of an empty capsid, as mutations which prevent capsid assembly or maturation do not cleave the newly replicated concatameric viral DNA [reviewed in L. A. Tengelsen et al, J. Virol., 67:3470-3480 (1993)]. Although it is known which cis-acting sequences in the viral DNA are required for cleavage and packaging, the viral proteins responsible for this cleavage/packaging event and their exact functions in this event have not been well characterized. Most of the information regarding the genes involved in this process has come from studies using temperature sensitive or engineered mutations in HSV-1. Along with the six genes required for capsid assembly (UL18, UL19, UL26, UL26.5, UL35, and UL38) reviewed in D. R. Thomsen et al, J. Virol., 68: 2442-2457 (1994), six other viral genes, UL6 [C. Addison et al, <u>Virol.</u>, <u>138</u>:246-259 (1984)], UL15 [C. Addison et al, <u>J.</u> Gen. Virol., 71:2377-2384 (1990)], UL25 [M.F. Al-Kobaisi et al, Virol., 180:380-388 (1991)], UL28 [G. Sherman and S. L. Bachenheimer, Virol., 158:427-434 (1987)], UL32 [G. Sherman and S.L. Bachenheimer, Virol., 163:471-480 (1988)], and UL33 [Y. Chang, et al., J. Virol., 70:3938-3946 (1996)], have been shown to be essential for DNA cleavage and packaging.

The genomic sequences for UL15 and UL19, as well as the sequences for the UL15 and UL19 gene products are available from GenBank under Accession No. 14112 (D00317). Further, the UL15 and UL19 sequences have been published [J. Baines et al, <u>J. Virol.</u>, <u>68</u>(12):8118-8124 (Dec. 1994) and Perry and Mcgeoch, <u>J. Gen. Virol.</u>, <u>69</u>:2831-2846 (1988), respectively]. The protein product of UL19 is VP5, also known as the major capsid protein.

Current methods for treating herpes virus infections have proved inadequate to prevent recurrence and viral shedding. Further, several of these treatments are toxic and are minimally effective.

What is needed are methods of identifying agents useful in treating and preventing infection with members of the herpesvirus family.

Summary of the Invention

In one aspect, the present invention provides a method for identifying an antiviral agent useful in treating infection with herpes viruses. The method involves allowing HSV-1 UL15 or a functional derivative or homologue thereof to come into association with a test compound. HSV-1 VP5 or a functional derivative or homologue thereof is then allowed to come into association with the HSV-1 UL15 or its derivative or homologue. Inhibition of the interaction between UL15 or its derivative or homologue and VP5 or its derivative or homologue is then determined, wherein said inhibition is indicative that the test compound is an antiviral agent.

In another aspect, the present invention provides a method for identifying an antiviral agent useful in treating infection with herpes viruses. The method involves allowing HSV-1 VP5 or a functional derivative or homologue thereof to come into association with a test compound. HSV-1 UL15 or a functional derivative or homologue thereof is then allowed to come into association with the HSV-1 VP5 or its derivative or homologue. Inhibition of the interaction between VP5 or its derivative or homologue and UL15 or its derivative or homologue is then determined, wherein said inhibition is indicative that the test compound is an antiviral agent.

In yet another aspect, the present invention provides a method for identifying antiviral agents which inhibit the function of the UL15/VP5 complex. This method involves allowing a sample containing a test compound to come into association with a herpes-infected cell and screening for viral inhibition. A test compound so identified is then screened

for the ability to block the function of a complex comprising UL15 or a functional derivative of cologue thereof and VP5 or a functional derivative or homologue thereof.

In a further aspect, the present invention provides compounds identified by the above methods.

Other aspects and advantages of the present invention are described further in the following detailed description of the preferred embodiments thereof.

Detailed Description of the Invention

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The present invention provides a method for identifying antiviral agents, particularly agents useful in preventing and treating infection with members of the herpes family, and compounds identified by the method of the invention. The method of the invention involves identifying molecules which prevent UL15/VP5 interaction and/or function.

For convenience, the amino acid sequences of UL15 and VP5 as provided in GenBank are reproduced in SEQID NO: 1 and SEQID NO:2, respectively. However, the present invention is not limited to the use of these amino acids sequences, as described below.

BINDING MOLECULE, as used herein, refers to molecules or ions which bind or interact specifically with UL15 protein or functional derivatives or homologues thereof, and/or bind or interact with the complex formed by UL15/VP5 (or functional derivatives or homologues thereof). Such binding molecules include, for example, small organic molecules, peptides, polypeptides, antibodies, particularly intracellular antibodies, antibody proteins and other antibody-derived reagents, chemical agents, nucleic acid sequences, enzyme substrates, cell membrane components and classical receptors. Interaction between the UL15 and/or UL19 gene products and such binding molecules, is indicative of anti-viral activity of the binding molecule. Prior to identification of a molecule as a binding molecule through the method of the invention, the molecule is referred to as a test compound.

As used herein, the term "FUNCTIONAL DERIVATIVE" of UL15 or VP5 includes fragments of these proteins, including peptides. Desirably, a UL15 peptide is characterized by the ability to form a complex with VP5 or a functional derivative or homologue thereof (e.g., by binding or otherwise interacting), and a VP5 peptide is characterized by the ability to form a complex with UL15 or a functional derivative or homologue thereof. For example, suitable fragments include, but are not limited to, fragments of UL15 located between about amino acid residue 1 to about amino acid residue 409 of SEQ ID NO:1, and between about residues 1 to about residues 508 of UL15 of SEQ ID NO:1. Other suitable UL15 proteins and peptides include those encoded by other splice variants of the UL15 gene. Similarly, included within this definition are homologues and/or peptides (i.e., fragments) of VP5. Functional derivatives also include UL15 and VP5 proteins and peptides which have been fused to or otherwise provided with a tag, e.g., an epitope tag, a polyHis tag, or the like, or a selectable marker or other detection means, for use in the various methods described herein. Suitable tags, markers and other means of detection are well known and can be readily selected by one of skill in the art.

Further included within the definition of functional derivatives are fusion proteins containing these UL15 or VP5 proteins or, preferably, peptides fused directly or indirectly (i.e. via linkers) to a fusion partner. Linkers and fusion partners for the peptides may be chosen for convenience in expression and/or purification. Suitable linkers may be readily selected by one of skill in the art. Similarly, suitable fusion partners, e.g., GST, Ga14, lexA, maltose-binding protein (MBP), lacZ, trp, and the like, may be readily selected by one of skill in the art. Also included within the definition of functional derivatives are UL15 and VP5 proteins which have been modified for ease of expression and/or purification.

For convenience throughout this application, the terms UL15 and VP5 will be used. However, it should be understood that one could readily substitute a functional derivative or homologue of either or both of these proteins in the methods of the invention.

Without wishing to be bound by theory, the inventors have found that the UL15 gene product specifically interacts with the major capsid protein VP5 (the gene product of UL19). As shown herein, experiments employing UL15 and VP5 proteins expressed using recombinant baculoviruses show that these two viral proteins are sufficient for this interaction. Furthermore, sequences contained within the amino terminus of UL15 appear to be critical for this interaction. This discovery, coupled with the knowledge that the UL15 gene and its homologues are the most highly conserved amongst all members of the *Herpesviridae*, permitted the inventors to develop the method of the invention. For convenience, the specification will refer to UL19 and VP5. However, as discussed, the methods described herein may be readily performed with functional derivatives or homologues of HSV-1 UL15 in other human herpesviruses, which include, but are not limited to, ORF42/45 in VZV, UL89 in HCMV, BD/BGRF1 in EBV, and NCBIgi 325496 in HHV-6, or suitable non-human herpesviruses. Similar methods can be performed with functional derivatives of UL19 or UL19 homologues derived from human or suitable non-human herpesviruses.

Thus, the present invention provides methods for screening and identifying agents which are active as anti-viral compounds for members of herpesviridae. Because UL15 and VP5 and their homologues are the most highly conserved proteins among the herpesvirus family, it is anticipated that an anti-viral agent identified by the method of the invention

will be useful in prevention and treatment of a, b and g herpes viruses. However, any select ti-viral agent of the invention may exhibit greater activity against viruses in one of these subgroups in comparison to its activity against viruses from another subgroup.

Desirably, the method of the invention involves the use of assays which detect or measure protein-protein associations. Particularly, the assays detect a molecule which binds to and occupies the binding site of UL15 thereby preventing binding to VP5, such that normal viral activity is prevented. Similarly, assays which detect a molecule which binds to and occupies the binding site of VP5 thereby preventing binding to UL15 and prevent normal viral activity, can be used. In another alternative, an assay may identify a molecule which inhibits the function of a UL15/VP5 complex. In yet another alternative, the assays may detect a molecule which binds to a site on UL15 or VP5 different from the normal binding site, which nevertheless inhibits the ability of UL15 and VP5 to interact and function normally, via allosteric effect.

Particularly, test compounds which interfere with the interaction between UL15 and VP5, or interfere with their function, have been found by the inventors to be useful as anti-viral agents for treatment and prophylaxis of herpes family viral infections. Most desirably, the test compound interacts with UL15 (or VP5) and inhibits its interaction with VP5 (or UL15). Thus, UL15 and VP5 can be used to identify and assess the binding of small molecule substrates and ligands in, for example, cells, cell-free preparations, chemical libraries, and natural product mixtures. These substrates and ligands may be natural substrates and ligands; may be structural or functional mimetics; or may be unrelated to the natural binding events. For example, the method of the invention may involve employing a suitable computer program to determine compounds having structures complementary to that of UL15, VP5, or portions thereof and screening those compounds for competitive binding to the protein.

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To identify small molecule antagonists, a rapid and reproducible assay is desirable for the screening of synthetic compounds, natural products, and peptides. Such an assay is desirably a protein based assay which would isolate UL15 and/or VP5 from interference by other assay components, particularly when utilizing cell-based assays. Desirably, the assay is adaptable to automation. Such assays are well known to those of skill in the art and are not intended to be a limitation on the present invention.

Where desired, and depending on the assay selected, UL15 (or, in an alternative embodiment, VP5) may be immobilized on a suitable surface. Such immobilization surfaces are well known. For example, a wettable inert bead may be used in order to facilitate multivalent interaction with UL15 (or VP5). Alternatively, an antibody to UL15 (or VP5) may be used. Typically, the surface containing UL15 (or VP5) is permitted to come into contact with a solution containing the test compound and a solution containing VP5 (or UL15) and binding is measured using an appropriate detection system. Suitable detection systems include the streptavidin horse radish peroxidase conjugate, direct conjugation by a tag, e.g., fluorescein. Other systems are well known to those of skill in the art. This invention is not limited by the detection system used.

Further, the methods of the invention are readily adaptable to combinatorial technology, where multiple molecules are contained on an immobilized support system. In such a method, a solution containing UL15 (or, in an alternative embodiment, VP5) is provided to the support system, followed by a solution containing VP5 (or UL15). Thus, the method of the invention permits screening of chemical compound and peptide based libraries.

The assay methods described herein are useful in screening for inhibition of the interaction between UL15 and VP5. In a preferred method, a solution containing the suspected inhibitors (i.e., the test compounds) is contacted with immobilized UL15 (or, in an alternative embodiment, VP5) substantially simultaneously with contacting the immobilized UL15 (or immobilized VP5) with the solution containing VP5 (or UL15). The solution containing the inhibitors may be obtained from any appropriate source, including, for example, extracts of supernatants from culture of bioorganisms, extracts from organisms collected from natural sources, chemical compounds, and mixtures thereof. In another variation, the inhibitor solution may be added prior to or after addition of VP5 (or UL15) to the immobilized UL15 (or VP5). As stated above, similar methods may be performed using homologues to UL15 or VP5, such as those identified above.

One example of a suitable assay is the yeast two-hybrid system. The yeast two-hybrid system provides methods for detecting the interaction between a first test protein (i.e., UL15 or vice-versa) and a second test protein (i.e., VP5 or vice-versa) using reconstitution of the activity of a transcriptional activator. The method is disclosed in U.S. Patent No.5,283,173; reagents are available from Clontech and Stratagene. Briefly, UL15 (or UL19) cDNA is fused to a Ga14 transcription factor DNA binding domain and expressed in yeast cells. Conversely, UL15 (or UL19) is fused to a transactivation domain of Ga14. Interaction of the fusion proteins via the UL15 and VP5 domains will lead to reconstitution of Ga14 activity and transactivation of expression of a reporter gene such as Ga11-lacZ.

As another example of a suitable assay, an enzyme-linked immunoassay (ELISA) or another immunoassay format may be utilized in which UL15 (or, in an alternative embodiment, VP5) is immobilized, directly or indirectly to a suitable surface. Such a immunoassay will typically involve use of specific antibodies (e.g., anti-UL15 or anti-VP5) or incubation of radiolabeled UL15 or VP5 with filter separation steps. In the presence of test substances which interrupt or inhibit formation of a UL15/VP5 complex, an increased amount of free UL15 or VP5 will be determined relative to a control lacking the test substance.

Yet another alternative method involves screening of peptide libraries for binding partner ecombinant tagged or labeled UL15 (or, in an alternative embodiment, VP5) is used to select peptides from a peptide or phosphopeptide library which interact with the UL15 (or, in an alternative embodiment, VP5). Sequencing of the peptides leads to identification of consensus peptide sequences which might be found in interacting proteins.

Another method is immunoaffinity purification. Recombinant UL15 (or, alternatively VP5) is incubated with labeled or unlabeled cell extracts and immunoprecipitated with anti-UL15 (or anti-VP5) antibodies. The immunoprecipitate is recovered with protein A-Sepharose and analyzed by SDS-PAGE. Unlabelled proteins are labeled by biotinylation and detected on SDS gels with streptavidin. The binding molecules are then analyzed for inhibition of the UL15/VP5 interaction.

This assessment of anti-viral activity of the test compounds identified as binding molecules can be performed using standard assays. Such assays will be readily apparent to one of skill in the art. Examples of such assays include, without limitation, plaque reduction assays and virus yield reduction assays.

In yet another aspect of the invention, methods for identifying antiviral agents by screening for inhibition of the function of the UL15/VP5 complex are provided. Suitable assays for performance of these methods are well known to those of skill in the art. For example, a cell-free system containing the UL15/VP5 complex may be utilized.

More desirably, however, cell-based assays are utilized in these screens. For example, the host cells may be modified to transiently or stably express UL15 and/or VP5. Alternatively, in a currently preferred embodiment, such an assay utilizes a cell infected with a herpes virus. A sample containing one or more test compounds is then brought into association with the infected cell and inhibition of normal herpes viral function is detected using methods such as those described herein and which are well known in the art. The test compound(s) identified are then screened for the ability to inhibit a complex comprising UL15 and VP5. This inhibition may be indirect, e.g., the compound may inhibit the function of the complex, or direct, e.g., by blocking binding of UL15 and VP5.

Test compounds identified by the methods described herein, which directly or indirectly (e.g., functionally) inhibit UL15/VP5 may be assayed using standard techniques.

Compositions

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The present invention further provides compounds identified by the methods of the invention and the use of the compounds as anti-viral agents useful in pharmaceutical compositions for the prophylaxis or treatment of infection with a member of the herpesvirus family.

The anti-viral agents identified by the method of the invention may be formulated into pharmaceutical compositions which may be used prophylactically or therapeutically, and preferably post-infection, to treat a herpes viral infection. The amount employed of the binding molecule/anti-viral agent will vary with the manner of administration, the employment of other active compounds, and the like, generally being in the range of about 1 mg to 10 mg, formulated in a solution of about 0.5 ml to 5 ml of a suitable carrier. Thus, the anti-viral agents of the present invention may be employed in combination with a non-sterile or sterile carrier or carriers for use with cells, tissues or organisms, such as a pharmaceutical carrier suitable for administration to a subject. Such compositions comprise, for instance, a media additive or a therapeutically effective amount of an anti-viral agent of the invention and a pharmaceutically acceptable carrier or excipient. Such carriers may include, but are not limited to, saline, buffered saline, dextrose, water, glycerol, ethanol and combinations thereof. The formulation should suit the mode of administration.

The pharmaceutical compositions may be administered in any effective, convenient manner including, for instance, administration by topical, oral, anal, vaginal, intravenous, intraperitoneal, intramuscular, subcutaneous, intranasal or intradermal routes, among others.

These examples are illustrative only and do not limit the scope of the invention.

Example 1 - Detection of UL15 Gene Product/VP5 Complex

UL15 was immunoprecipitated from HSV-1 infected Vero cells [ATCC, CCL81], and the immunoprecipitate was immunoblotted with antisera to the various capsid proteins. More particularly, Vero cells were infected with 10 PFU per cell of HSV-1 (KOS) [K. O. Smith, <u>Proc. Soc. Exp. Biol. Med., 115</u>:814-816 (1964); P. A. Schaffer et al, <u>Virology, 52</u>: 57-71 (1973)] and were harvested 20 hours post infection. Cells were lysed by freeze-thawing three times in phosphate buffered saline (PBS) followed by centrifugation to pellet debris. The supernatant was pre-cleared with normal rabbit serum and protein-A bound to sepharose. The resulting supernatant was immunoprecipitated with five microliters of UL15-specific antisera (NC-1) [J. D. Baines, <u>J. Virol., 68</u>(12):8118-8124 (December 1994)], immunoblotted, and probed with NC-1 antisera to the HSV-1 capsid proteins [G. H. Cohen et al, <u>J. Virol., 34</u>(2):521-531 (May 1980)].

The UL15 polyclonal antisera directed against the carboxy-terminal portion of the UL15 protein that was used has been previously shown to react with two proteins of 75 kD and 35 kD molecular weight [Baines et al, cited above]. Immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates of UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates are under the UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates are under the UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates are under the UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates are under the UL15 from HSV-1 infected Vero cells were also found to contain a 155 kD protein which immunoprecipitates are under the UL15 from HSV-

noreacts with antisera against the major capsid protein VP5 and corresponds with the report. De of VP5. This band was not present in uninfected Vero cells. Conversely, a band of 75 kD which was immunoreactive with antisera against UL15 was detected when VP5 was immuno-precipitated from HSV-1 infected Vero cells, but not uninfected Vero cells. Furthermore, none ofthe 35 kD UL15 protein was detected in the VP5 immunoprecipitates, nor was any UL15 5 protein detected in a preparation of sucrose-density banded, purified HSV-1 capsids, suggesting that UL15 is not a capsid protein. Probing of immunoblots of UL15 immuno-precipitates with antisera to the other HSV-1 capsid proteins did not reveal co-immuno-precipitation with these other proteins. This could be due to either a weak interaction which was not detectable by this method, or by the low abundance of the capsid protein in the UL15 immunoprecipitate, as several capsid genes, such as VP21 and VP24, are not present in great amounts within the capsid. The data indicates that in HSV-1 infected cell extracts, UL15 and VP5 complex together.

Example 2 - Characterization of UL15/VP5 Complex

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To determine if the UL15/VP5-containing complex identified in Example 1 contains any other proteins, whether UL15 and VP5 directly interact within the complex, or whether only UL15 and VP5 are sufficient for complex formation, the following study was performed.

UL15 and VP5 were expressed recombinantly using a baculovirus expression system. The open reading frames (ORFs) of the UL15 (VP5) gene was derived as partial cDNA from Vero cells infected form HSV-1 (KOS) and pSG10 and pSG16 plasmids containing the gene [described in A. L. Goldin et al, J. Virol., 38(10):50-58 (April 1981)]. The UL19 ORF was similarly derived, using plasmid pSG16 [Goldin et al, cited above]. The UL15 and UL19 ORFs were cloned into the baculovirus expression vector pVL 1393 [Pharminogen] and used to generate recombinant baculoviruses. Susceptible Sf9 cells [ATCC] were co-infected with these two recombinant baculoviruses (BAC-UL 15 and BAC-VP5) at a multiplicity-of-infection (MOI) of 5 PFU of each virus per cell. Total protein lysates were prepared at 48 hours by direct lysis in I mI of sample buffer. Uninfected Sf9 cells, cells infected wild type baculovirus, cells infected with wild-type baculovirus and BAC-UL15, and cells infected with wild type baculovirus and BAC-VP5 were used as controls. Twenty-five microliters of this sample was loaded onto a 4-20% gradient polyacrylamide gel containing 0.1% SDS, and electrophoresed in Tris-glycine buffer. Following electrophoresis and electrotransfer, the gel was immunoblotted and probed with rabbit polyclonal antisera specific for either VP5 (NC1) (using UL15 antisera) or UL15 (using VP5 antisera).

Specifically immunoreactive bands of the appropriate sizes (75 kD for UL15) and (155 kD for VP5) were observed in cells infected with the respective baculovirus and not in uninfected Sf9 cells or Sf9 cells infected with wild type baculovirus.

Example 3 - Specificity of UL15:VP5 Complex

Because VP5 is a rather large protein (155 kD), it was important to determine if UL15:VP5 complex formation was due to a non-specific sticking of the two proteins. To address this question, UL15 and several other unrelated polypeptides were synthesized *in vitro* by coupled transcription/translation and tested for their ability to interact with VP5 expressed by baculovirus as follows.

Template DNAs containing the open reading frames of UL15, and negative controls HSV-1 UL12 (alkaline exonuclease), human papillomavirus (HPV) type 6b E1 (HPV-6b E1), HPV-16 E1, HPV-18 E2, and HPV-16 E2 proteins were used to synthesize their respective radiolabeled proteins in vitro in a 50 microliter synthesis reaction. A small portion of the reaction (1/10th) was analyzed by SDS-PAGE to confirm the expression of these proteins. To determine if VP5 interacted with these proteins, the remainder of the reaction was mixed with 50 ml of an extract made from BAC-VP5 infected Sf9 cells, immunoprecipitated with VP5 antisera, and analyzed by SDS-PAGE.

Although a background amount of HPV-6b E1, HPV-16 E1, and HPV-16 E2 proteins co-immunoprecipitated with VP5, approximately a 10-fold greater amount of UL15 was co-immunoprecipitated with VP5, strongly suggesting that the interaction of UL15 and VP5 was due to specific interaction with VP5.

Example 4 - Identification of Region of UL15 Interacting with VP5

To determine which region of UL15 was involved in complex formation with VP5, a series of UL15 polypeptides which were truncated at the carboxy-terminus were synthesized *in vitro* by coupled transcription/translation and examined for their ability to interact with VP5.

Truncated UL15 proteins which contained amino acids 409-735 [SEQ ID NO:1] were still able to coimmunoprecipitate with VP5. However, UL15 proteins which further deleted amino acids between 1 - 409 [SEQ ID NO: 1] were unable to interact with VP5, suggesting that amino acids in the amino-terminus of UL15 are critical for VP5 interaction. This finding is also confirmed by the observation that the 35kD form of UL15 which lacks the amino-terminal of the 75 kD protein, but shares only the carboxy-terminal region does not coimmunoprecipitate with VP5.

The data presented in Examples 1-4 show that the 75 kD protein from the UL15 gene communoprecipitates with the major capsid protein VP5. This interaction may provide a means for linking the terminase complex with the empty capsid so that the newly replicated DNA can be easily packaged. Inspection of the UL15 sequence reveals the presence of a potential cleavage site for the UL26 protease located at amino acids 332-339 [SEQ ID NO:1]. If this site represents a genuine substrate for the UL26 protease, it may explain the generation of the 35 kD protein, and also provide a means by which UL15 terminase activity could be inactivated signalling the completion of DNA packaging.

Example 5 - Application of UL15 and VP5 proteins in a screen for molecules blocking the interaction of UL15 with VP5

The following assay format may be used to identify molecules which inhibit UL15:VP5 binding by screening a large bank of chemical compounds and natural products.

The VP5 protein is biotinylated for simplicity of assay and for ease of detection. Biotinylation is carried out essentially as described in <u>Avidin-Biotin Chemistry</u>: A handbook, M. D. Savage *et al.*, Pierce Chemical Company (1992). All steps of the assay after coating are carried out at room temperature.

The wells of 96 well microtiter plates (Immunlon 4, Dynatech Laboratories) are coated with UL15 5 gene product (2 mg/ml) in 100 ml/well of 0.1 M sodium bicarbonate, pH 9.4 and incubated overnight @ 4°C. Immediately afterwards, dilutions of test compound are added (10 ml). The compounds are dissolved at 100x assay concentration in dimethyl sulfoxide (DMSO) and subsequently diluted in 50%DMSO/50% H₂O to a 10X working stock.

The wells are washed with PBS (phosphate buffered saline) and blocked with 0.5% gelatin in PBS for 1 hour. Following an additional PBS wash, biotinylated VP5 is serially diluted in PBS containing 1 mg/ml BSA, 0.05% Tween directly in the wells in a final volume of 0.1 ml and incubated for 1 hour. The wells are washed with PBS and bound VP5 protein is measured by the addition of 0.1 ml of strepavidin-HRP (streptavidin conjugated with horseradish peroxidase (Southern Biotech)) at a 1:2000 dilution for 1 hour, followed by washing and color development with 100 ml ABTS substrate (Kierkegaard and Perry Laboratories Inc., Maryland) and measurement of absorbance at 405 nm. In some cases the color reactions are arrested by addition of 100 ml of 1% SDS prior to measurement of absorbance.

Numerous modifications and variations of the present invention are included in the above-identified specification and are expected to be obvious to one of skill in the art. Such modifications and alterations to the compositions and processes of the present invention are believed to be encompassed in the scope of the claims appended hereto.

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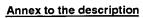
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SEQUENCE LISTING

5		
	(1) GENE	RAL INFORMATION:
10	(i)	APPLICANT: SmithKline Beecham Corporation
	(ii)	TITLE OF INVENTION: Use of HSV-1 UL-15 and VP5 in Identifying Anti-Viral Agents
15	(iii)	NUM EER OF SEQUENCES: 2
	(iv)	CORRESPONDENCE ADDRESS: (A) ADDRESSEE: F J Cleveland & Company (B) STREET: 40/43 Chancery Lane
20		(C) CITY: London (D) COUNTY: (E) COUNTRY: United Kingdom (F) POST CODE: WC2A 1JQ
25	(v)	COMPUTER READABLE FORM: (A) MEDIUM TYPE: Floppy disk (B) COMPUTER IEM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS (D) SOFTWARE: Patentin Release #1.0, Version #1.30
30	(vi)	CURRENT APPLICATION DATA: (A) APPLICATION NUMBER US (B) FILING DATE: (C) CLASSIFICATION:
35	(viii)	ATTORNEY/AGENT INFORMATION: (A) NAME: CRUMP, Julian Richard John (B) GENERAL AUTHORISATION NUMBER 37127
40	(ix)	(C) REFERENCE/DOCKET NUMER P50617 TELECOMMUNICATION INFORMATION: (A) TELEPHONE: +44 171 405 5875 (B) TELEFAX: +44 171 831 0749
45	(2) INFO	RMATION FOR SEQ ID NO: 1:
50	(i)	SEQUENCE CHARACTERISTICS: (A) LENGTH: 735 amino acids (B) TYPE: amino acid (C) STRANDEDNESS: (D) TOPOLOGY: unknown
55	(ii)	MOLECULE TYPE: protein

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

	Met 1	Phe	Gly	Gln	Gln 5	Leu	Ala	Ser	Asp	Val 10	Gln	Gln	Tyr	Leu	Glu 15	Arg
5	Leu	Glu	Lys	Gln 20	Arg	Gln	Leu	Lys	Va] 25	Gly	Ala	Asp	Glu	Ala 30	Ser	Ala
10	Gly	Leu	Thr 35	Met	Gly	Gly	Asp	Ala 40	Leu	Arg	Val	Pro	Phe 45	Leu	Asp	Phe
	Ala	Thr 50	Ala	Thr	Pro	Lys	Arg 55	His	Gln	Thr	Val	Val 60	Pro	Gly	Val	Gly
15	Thr 65	Leu	His	Asp	Cys	Cys 70	Glu	His	Ser	Pro	Leu 75	Phe	Ser	Ala	Val	Ala 80
	Arg	Arg	Leu	Leu	Phe 85	Asn	Ser	Leu	Val	Pro 90	Ala	Gln	Leu	Lys	Gly 95	Arg
20	Asp	Phe	Gly	Gly 100	Asp	His	Thr	Ala	Lys 105	Leu	Glu	Phe	Leu	Ala 110	Pro	Glu
25		Val	115					120			_		1 25			
		Val 130					135					140				
30	145	Ala				150					155					160
		Asp			165					170					175	
35		Glu		180					185					190		
40		His Met	195					200					205	-		
		210 Glu					215					220				
45	225	Ser				230					235					240
50		Val			245					25 0					255	
		Leu		260			_		265					270		
55		Ile	275					280					285			
	114.0	110	1129	دود	, <u></u> u	1111	JIU	0	ACIT	T 11 G	JLU	G L U	116	ν»h	n.a	cys

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290 295 300 Leu Arg Gly Trp Phe Gly Ser Ala Arg Val Asp His Val Lys Gly Glu 310 5 315 Thr Ile Ser Phe Ser Phe Pro Asp Gly Ser Arg Ser Thr Ile Val Phe 330 Ala Ser Ser His Asn Thr Asn Gly Ile Arg Gly Gln Asp Phe Asn Leu 10 345 Leu Phe Val Asp Glu Ala Asn Phe Ile Arg Fro Asp Ala Val Gln Thr 360 15 Ile Met Gly Phe Leu Asn Gln Ala Asn Cys Lys Ile Ile Phe Val Ser 375 Ser Thr Asn Thr Gly Lys Ala Ser Thr Ser Phe Leu Tyr Asn Leu Arg 20 390 3 95 Gly Ala Ala Asp Glu Leu Leu Asn Val Val Thr Tyr Ile Cys Asp Asp His Met Pro Arg Val Val Thr His Thr Asn Ala Thr Ala Cys Ser Cys 25 4 25 Tyr Ile Leu Asn Lys Pro Val Phe Ile Thr Met Asp Gly Ala Val Arg 440 30 Arg Thr Ala Asp Leu Phe Leu Ala Asp Ser Phe Met Gln Glu Ile Ile 455 Gly Gly Gln Ala Arg Glu Thr Gly Asp Asp Arg Pro Val Leu Thr Lys 35 Ser Ala Gly Glu Arg Phe Leu Leu Tyr Arg Pro Ser Thr Thr Thr Asn 490 Ser Gly Leu Met Ala Pro Asp Leu Tyr Val Tyr Val Asp Pro Ala Phe 40 Thr Ala Asn Thr Arg Ala Ser Gly Thr Gly Val Ala Val Val Gly Arg 5.20 45 Tyr Arg Asp Asp Tyr Ile Ile Phe Ala Leu Glu His Phe Phe Leu Arg 530 535

Arg Val Ala Val Glu Gly Asn Ser Ser Gln Asp Ser Ala Val Ala Ile
580 . 585 . 590

Ala Leu Thr Gly Ser Ala Pro Ala Asp Ile Ala Arg Cys Val Val His

Ser Lou Thr Gln Val Leu Ala Leu His Pro Gly Ala Phe Arg Gly Val

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		Ala	Thr	His 595	Val	His	Thr	Glu	Met 600	His	Arg	Leu	Leu	Ala 605	Ser	Glu	Gly
5		Ala	Asp 610	Ala	Gly	Ser	Gly	Pro 615	Glu	Leu	Leu	Phe	Tyr 620	His	Cys	Glu	Pro
10		Pro 625	Gly	Ser	Ala	Val	Leu 630	Tyr	Pro	Phe	Phe	Leu 635	Leu	Asn	Lys	Gln	Lys 640
		Thr	Pro	Ala	Phe	Glu 645	His	Phe	Ile	Lys	Lys 650	Phe	Asn	Ser	Gly	Gly 655	Val
15		Met	Ala	Ser	Gln 660	Glu	Ile	Val	Ser	Ala 665	Thr	Val	Arg	Leu	Gln 670	Thr	Asp
20		Pro	Val	Glu 675	Туr	Leu	Leu	Glu	Gln 680	Leu	Asn	Asn	Leu	Thr 685	Glu	Thr	Val
		Ser	Pro 690	Asn	Thr	Asp	Val	Arg 695	Thr	Tyr	Ser	Gly	Lys 700	Arg	Asn	Gly	Ala
25		Ser 705	Asp	Asp	Leu	Met	Val 710	Ala	Val	Ile	Met	Ala 715	Ile	Tyr	Leu	Ala	Ala 720
		Gln	Ala	Gly	Pro	Pro 725	His	Thr	Phe	Ala	Pro 730	Ile	Thr	Arg	Val	Ser 735	
30																	
35	(2)	INFO	RMAT I	ION I	FOR :	SEQ I	ID NO	D: 2:									
40		(i)	(A) (B) (C)	LEI TYI ST:	E CHANGTH: PE: a RANDE	137 amino EDNE:	74 ar o aci SS:	mino id		ds							
45		(ii)	MOLE	ECULE	E TYE	PE: p	orote	ei n									
		(xi)	SEQU	JENC	E DES	SCRIE	PTIO	N: SE	EQ ID	NO:	2:						
50		Met 1	Ala	Ala	Pro	As n 5	Arg	Asp	Pro	Pro	Gly 10	Туr	Arg	Tyr	Ala	Ala 15	Ala
		Met	Val	Pro	Thr 20	Gly	Ser	Leu	Leu	Ser 25	Thr	Ile	Glu	Val	Ala 30	Ser	His
55		Arg	Arg	Leu 35	Phe	Asp	Phe	Phe	Ser 40	Arg	Val	Arg	Ser	Asp 45	ALa	Asn	Ser

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	Leu	Tyr 50	Asp	Val	Glu	Phe	Asp 55	Ala	Leu	Leu	Gly	Ser 60	Tyr	Cys	Asn	Thr
5	Leu 65	Ser	Leu	Val	Arg	Phe 70	Leu	Glu	Leu	Gly	Leu 75	Ser	Val	Ala	Cys	Val 80
	Cys	Thr	Lys	Phe	Pro 85	Glu	Leu	Ala	Tyr	Met 90	Asn	Glu	Gly	Arg	Val 95	Gln
10	Phe	Glu	Val	His 100	Gln	Pro	Leu	Ile	Ala 105	Arg	Λsр	Gly	Pro	His 110	Pro	Ile
15	Glu	Gln	Pro 115	Thr	His	Asn	Tyr	Met 120	Thr	Lys	Ile	Ile	As p 1 25	Arg	Arg	Ala
	Leu	As n 130	Ala	Ala	Phe	Ser	Leu 135	Ala	Thr	Glu	Ala	Ile 140	Ala	Leu	Leu	Thr
20	Gly 145	Glu	Ala	Leu	Asp	Gly 150	Thr	Gly	Ile	Gly	Ala 155	His	Arg	Gln	Leu	Arg 160
25	Ala	Ile	Gln	Gln	Leu 165	Ala	Arg	Asn	Val	Gln 170	Ala	Val	Leu	Gly	Ala 175	Phe
20	Glu	Arg	Gly	Thr 180	Ala	Asp	Gln	Met	Leu 185	His	Val	Leu	Leu	Glu 190	Lys	Ala
30	Pro	Pro	Leu 195	Ala	Leu	Leu	Leu	Pro 200	Met	Gln	Arg	Tyr	Leu 205	Asp	Asn	Gly
	Arg	Leu 210	Ala	Thr	Arg	Val	Ala 215	Arg	Ala	Thr	Leu	Val 220	Ala	Glu	Leu	Lys
35	Arg 225	Ser	Phe	Cys	Glu	Thr 230	Ser	Phe	Phe	Leu	Gly 235	Lys	Ala	Gly	His	Arg 240
40	Λrg	Glu	Ala	Val	Glu 245	Ala	Trp	Leu	Val	As p 25 0	Leu	Thr	Thr	Ala	Thr 255	Gln'
	Pro	Ser	Val	Al a 260	Val	Pro	Arg	Leu	Thr 265	His	Ala	Asp	Thr	Arg 270	Cly	Arg
45	Pro	Val	Asp 275	Gly	Val	Leu	Val	Thr 280	Thr	Ala	Pro	Ile	Lys 285	Gln	Arg	Leu
	Leu	Gln 290	Ser	Phe	Leu	Lys	Val 295	Glu	Asp	Thr		Ala 300	Asp	Val	?ro	Val
50	Thr 305	Tyr	Gly	Glu		Val 310	Leu	Asn	Gly	Ala	Asn 315	Leu	Val	Thr	Ala	Leu 3 20
55	Val	Met	Gly	Lys	Ala 325	Val	Arg	Ser	Leu	Asp 330	Asp	Val	Gly	Arg	His 335	Leu
	Leu	Glu	Met	Gln	Glu	Glu	Gln	Leu	Asp	Leu	Asn	Arg	Gln	Thr	Leu	Asp





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5	Glu	Leu	Glu 355	Ser	Ala	Pro	Gln	Thr 360	Thr	Arg	Val	Arg	Ala 365	Asp	Leu	Val
	Ser	Ile 370	Gly	Glu	Lys	Leu	Val 375	Phe	Leu	Glu	Ala	Leu 380	Glu	Lys	Arg	Ile
10	Tyr 385	Ala	Ala	Thr	Asn	Val 390	Pro	Туr	Pro	Leu	Val 395	Gly	Ala	Met	Asp	Leu 400
15	Thr	Phe	Val	Leu	Pro 405	Leu	Gly	Leu	Phe	Asn 410	Pro	Val	Met	Glu	Arg 415	Phe
	Ala	Ala	His	Ala 420	Gly	Asp	Leu	Val	Pro 425	Ala	Pro	Gly	His	Pro 430	Asp	Pro
20	Arg	Ala	Phe 435	Pro	Pro	Arg	Gln	Leu 440	Phe	Phe	Trp	Gly	Lys 445	Asp	Arg	Gln
	Val	Leu 450	Arg	Leu	Ser	Leu	Glu 455	His	Ala	Ile	Gly	Thr 460	Val	Cys	His	Pro
25	Ser 465	Leu	Met	Asn	Val	Asp 470	Ala	Ala	Val	Gly	Gly 475	Leu	Asn	Arg	Asp	Pro 480
30	Val	Glu	Ala	Ala	As n 485	Pro	Tyr	Gly	Ala	Tyr 490	Val	Ala	Ala	Pro	Ala 495	Gly
	Pro	Ala	Ala	Asp 500	Met	Gln	Gln	Leu	Phe 505	Leu	Asn	Ala	Trp	Gly 510	Gln	Arg
35	Leu	Ala	His 515	Gly	Arg	Val	Arg	Trp 520	Val	Ala	Glu	Gly	Gl n 5 25	Met	Thr	Pro
	Glu	Gln 530	Phe	Met	Gln	Pro	As p 535	Asn	Ala	Asn	Leu	Ala 540	Leu	Glu	Leu	His
40	Pro 545	Ala	Phe	Asp	Phe	Phe 550	Val	Gly	Väl	Ala	Asp 555	Val	Glu	Leu	Pro	Gly 560
45	Gly	Asp	Val	Pro	Pro 565	Ala	Gly	Pro	Gly	Glu 570	Ile	Gln	Ala	Thr	T rp 575	Arg
	Val	Val	Asn	Gly 580	Asn	Leu	Pro	Leu	Ala 585	Leu	Cys	Pro	Ala	Ala 590	Phe	Arg
50	Asp	Ala	Arg 595	Gly	Leu	Glu	Leu	Gly 600	Val	Gly	Arg	His	Ala 605	Met	Ala	Pro
	Ala	Thr 610	Ile	A_a	Ala	Val	Arg 615	Gly	Ala	Phe	Asp	As p 6 20	Arg	Asn	Туг	Pro
55	Ala 625	Val	Phe	Tyr	Leu	Leu 630	Gln	Ala	Ala	Ile	His 635	Gly	Ser	Glu	His	Val 640

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	Phe	Cys	Ala	Leu	Ala 645	Arg	Leu	Val	Val	Gln 650	Cys	Ile	Thr	Ser	Tyr 655	Trp
5	Asn	Asn	Thr	Arg 660	Cys	Ala	Ala	Phe	Val 665	Asn	Asp	Tyr	Ser	Leu 670	Val	Ser
10	Туг	Val	Val 675	Thr	Tyr	Leu	Gly	Gly 680	Asp	Leu	Pro	Glu	Glu 685	Cys	Met	Ala
	Val	Tyr 690	Arg	Asp	Leu	Val	Ala 695	His	Val	Glu	Ala	Leu 700	Ala	Gln	Leu	Val
15	As p 705	Asp	Phe	Thr	Leu	Thr 710	Gly	Pro	Glu	Leu	Gly 715	Gly	Gln	Ala	Gln	Ala 720
00	Glu	Leu	Asn	His	Leu 7 25	Met	Arg	Asp	Pro	Ala 730	Leu	Leu	Pro	Pro	Leu 735	Val
20	Trp	Asp	Cys	Asp 740	Ala	Leu	Met	Arg	Arg 745	Ala	Ala	Leu	Asp	Arg 750	His	Arg
25	Asp	Cys	Arg 755	Val	Ser	Ala	Gly	Gly 760	His	Asp	Pro	Val	Tyr 765	Ala	Ala	Ala
	Cys	As n 770	Val	Ala	Thr	Ala	As p 775	Phe	Asn	Arg	Asn	Asp 780	Gly	Gln	Leu	Leu
30	His 785	Asn	Thr	Gln	Ala	Arg 790	Ala	Ala	Asp	Ala	Ala 795	Asp	Asp	Arg	Pro	His 800
35	Arg	Gly	Ala	Asp	Trp 805	Thr	Val	His	His	Lys 810	Ile	Tyr	Tyr	Туг	Val 815	Met
	Val	Pro	Ala	Phe 820	Ser	Arg	Gly	Arg	Cys 8 25	Cys	Thr	Ala	Gly	Val 830	Arg	Phe
40	Asp	Arg	Val 835	Tyr	Ala	Thr	Leu	Gln 840	Asn	Met	Val	Val	Pro 845	Glu	Ile	Ala
	Pro	Gly 850	Glu	Glu	Cys	Pro	Ser 855	Asp	Pro	Val	Thr	Asp 860	Pro	Ala	His	Pro
45	Leu 865	His	Pro	Ala	Asn	Leu 870	Val	Ala	Asn	Thr	Val 875	Asn	Ala	Met	Phe	His 880
50	Asn	Gly	Arg	Val	Val 885	Val	Asp	Gly	Pro	Ala 890	Met	Leu	Thr	Leu	Gln 895	Val
	Leu	Ala	His	As n 900	Met	Ala	Glu	Arg	Thr 905	Thr	Ala	Leu	Leu	Cys 910	Ser	Ala
55	Ala	Pro	Asp 915	Ala	Gly	Ala	Asn	Thr 920	Ala	Ser	Thr	Thr	As n 925	Met	Arg	Ile

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	Phe	As p 930	Gly	Al.a	Leu	His	Ala 935	Gly	Ile	Leu	Leu	Met 940	Ala	Pro	Gln	His
5	Leu 945	Asp	His	Thr	Ile	G1 n 95 0	Asn	Gly	Asp	Tyr	Phe 955	Tyr	Pro	Leu	Pro	Val 960
	His	Ala	Leu	Phe	Ala 965	Gly	Ala	Asp	His	Val 970	Ala	Asn	Ala	Pro	As n 975	Phe
10	Pro	Pro	Ala	Leu 980	Arg	Asp	Leu	Ser	Arg 985	Gln	Val	Pro	Leu	Val 990	Pro	Pro
15	Ala	Leu	Gly 995	Ala	Asn	Tyr	Phe	Ser 1000		Ile	Arg	Gln	Pro 1005		Val	Gln
	His	Val 1010		Glu	Ser	Ala	Ala 1015		Glu	Asn	Ala	Leu 1020		Tyr	Ala	Leu
20	Met 1025		GTA	Туг	Phe	Lys 1030		Ser	Pro	Val	Ala 1035		His	His	Gln	Leu 1040
	Lys	Thr	Gly	Leu	His 1045		Gly	Phe	Gly	Phe 1050		Val	Val	Arg	Gln 1055	-
25	Arg	Phe	Val	Thr 1060		Asn	Val	Leu	Phe 1065		Glu	Arg	Ala	Ser 1070		Ala
30	Туг	Phe	Leu 1075	Gly	Gln	Leu	Gln	Val 1080		Arg	His	Glu	Thr 1085		Gly	Gly
	Val	As n 1090		Thr	Leu	Thr	Gln 1095		Arg	Ala	Asn	Val 1100	_	Leu	Gly	Val
35	Gly 1105		Thr	Ala	Val	Val 1110		Thr	Ala	Thr	Val 1115	_	Asn	Pro	Val	Thr 1120
40	Asp	Met	Gly	Asn	Leu 1125		Gln	Asn	Phe	Tyr 1130		Gly	Arg	Gly	Ala 1135	
40	Pro	Leu	Leu	Asp 1140		Ala	Ala	Ala	Val 1145		Leu	Arg	Asn	Aia 1150		Val
45	Ala	Gly	Asn 1155	Arg	Leu	Gly	Pro	Ala 1160		Pro	Val	Pro	Val 1165		Gly	Cys
	Ala	Gln 1170		Pro	Arg	Arg	Ala 1175		Met	Asp	His	Gly 1180		Asp	Ala	Val
50	Cys 1185		Phe	Ile	Ala	Thr 1190		Val	Ser	Thr	Asp 1195		Asn	Tyr	Phe	Arg 1200
	Arg	Pro	Cys	Asn	Pro 1205		Gly	Arg	Ala	Ala 1210		Gly .	Val	Туr	Ala 1215	
55	Asp	Lys	Glu	Gly	Asp	Val	Thr	Ala	Leu	Met	Tyr	Asp	His	Gly	Gln	Ser



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5	Asp	Pro	Ser 1235		Ala	Phe	Ala	Ala 1240		Ala	Asn	Pro	Trp 1245		Ser	Gln
	Arg	Phe 1250		Tyr	Gly	Asp	Leu 1 25 5		Tyr	Asn	Gly	Ala 1260	Tyr)	His	Leu	Asn
10	Gly 1265		Ser	Pro	Val	Leu 1270		Pro	Cys	Phe	Lys 1275		Phe	Thr	Ser	Ala 1280
15	Asp	Ile	Ala	Ala	Lys 1 285		Arg	Cys	Leu	Glu 1290		Leu	Ile	Val	Glu 1295	
	Gly	Ser	Ala	Val 1300		Thr	Alā	Thr	Ala 1305		Ser	Asp	Val	Gln 1310		Lys
20	Arg	Pro	Pro 1315		Cys	Arg	Glu	Leu 1320		Glu	Asp	Pro	Cys 1325		Leu	Phe
	Gln	Glu 1330		Tyr	Pro	Leu	Thr 1335		Ala	Ser	Asp	Pro 1340	Ala)	Leu	Leu	Arg
25	Ser 1345		Arg	Asn	Gly	Glu 1350		His	Ala	Arg	Glu 1355		His	Phe	Ala	Gln 1360
30	Tyr	Leu	Val	Tyr	Asp 1365		Ser	Pro	Leu	Lys 1370		Leu	Ala	Leu		

Claims

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- 35 1. A method for identifying an antiviral agent useful in treating infection with herpes viruses,
 - (a) providing UL15 of herpes simplex virus-1 (HSV-1) or a functional derivative or homologue thereof;
 - (b) permitting UL15 or its derivative or homologue to come into association with a test compound;
 - (c) providing VP5 of HSV-1 or a functional derivative or homologue thereof;
 - (d) screening for inhibition of the interaction between UL15 or its derivative or homologue and VP5 or its derivative or homologue, wherein said inhibition is indicative that the test compound is an antiviral agent.
 - 2. A method for identifying an antiviral agent useful in treating infection with herpes viruses,
 - (a) providing VP5 of herpes simplex virus-1 (HSV-1) or a functional derivative or homologue thereof;
 - (b) permitting VP5 or its derivative or homologue to come into association with a test compound;
 - (c) providing UL15 of HSV-1 or a functional derivative or homologue thereof;
 - (d) screening for inhibition of the interaction between the VP5 or its derivative or homologue and UL15 or its derivative or homologue, wherein said inhibition is indicative that the test compound is an antiviral agent.
 - 3. The method according to claim 1 or 2 wherein a UL15/VP5 complex comprising UL15 or a functional derivative or homologue thereof and VP5 or a functional derivative or homologue thereof is permitted to come into association with a test compound.
- 55 4. The method according to claim 1 or 2 wherein the test compound binds the UL15/VP5 complex.
 - The method according to claim 1 or 2 wherein the test compound binds UL15 or a functional derivative or homologue thereof.

- 6. The method according to claim 1 or 2 wherein the test compound binds VP5 or a function and rivative or homologue thereof.
- 7. The method according to claim 1 or 2 wherein the functional derivative of UL15 is selected from the group consisting of:
 - (a) SEQ ID NO:1;

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- (b) a peptide from SEQ ID NO:1;
- (c) a fusion protein comprising (a) or (b) and a selected fusion partner;
- (d) a tagged protein comprising (a) or (b) and a selected marker or tag.
- 8. The method according to claim 7, wherein the UL15 peptide is selected from the group consisting of:
- (a) about amino acid 1 to about amino acid 409 of SEQ ID NO:1;and
 - (b) about amino acid 1 to about amino acid 508 of SEQ ID NO:1.
- The method according to claim 7, wherein the fusion partner is selected from the group consisting of: GST, Ga14,
 lexA, maltose-binding protein, lacZ, and Trp.
 - 10. The method according to claim 1 or 2, wherein the functional derivative of VP5 is selected from the group consisting of:
 - (a) SEQ ID NO:2;
 - (b) a peptide from SEQ ID NO:2;
 - (c) a fusion protein comprising (a) or (b) and a selected fusion partner.
 - (d) a tagged protein comprising (a) or (b) and a and a selected marker or epitope tag.
- 30 11. The method according to claim 1 or 2 wherein the UL15 homologue is selected from the group consisting of:

varicella-zoster virus ORF42/45; human cytomegalovirus UL89; epstein-barr virus BD/BGRF1; and human herpes virus 6 in NCBIgi 325496.

- 12. The method according to claim 1 or 2 wherein UL15 is immobilized on a solid support.
- 13. The method according to claim 1 or 2 wherein VP5 is immobilized on a solid support.
- 14. The method according to claim 1 or 2 wherein said screen is a yeast 2-hybrid screen.
- 15. The method according to claim 1 or 2 wherein said screen is an enzyme-linked immunosorbent assay.
- 45 16. The method according to claim 1 or 2 wherein the test compound is immobilized on a support system.
 - 17. The method according to claim 1 or 2, wherein said test compound is a chemical agent.
 - 18. The method according to claim 1 or 2, wherein said test compound is a polypeptide.
 - 19. The method according to claim 1 or 2, wherein said test compound is an intracellular antibody.
 - 20. The method according to claim 1 or 2 wherein steps (b) and (c) occur substantially simultaneously.
- 21. A compound identified by the method according to claim 1 or 2.
 - 22. A pharmaceutical composition comprising a compound according to claim 21 and a pharmaceutically acceptable carrier.

- 23. A method for identifying antiviral agents which functionally inhibit a UL15/VP5 complementary omprising UL15 or a functional derivative or homologue thereof, said method comprising the steps of:
 - (a) providing a cell infected with a herpes virus;

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- (b) permitting a sample containing a test compound to come into association with the herpes-infected cell;
- (c) identifying a test compound which inhibits the herpes virus;
- (d) screening the test compound which is identified in (c) for the ability to block the function of the UL15/VP5 complex.
- 24. A method for identifying antiviral agents which functionally inhibit a UL15/VP5 complex comprising UL15 or a functional derivative or homologue thereof, said method comprising the steps of:
 - (a) providing a cell-free system containing the UL15/VP5 complex;
 - (b) permitting a sample containing a test compound to come into association with the complex; and
 - (c) identifying a test compound which inhibits the function of the UL15/VP5 complex.
- 25. A method for identifying antiviral agents which functionally inhibit a UL15/VP5 complex comprising UL15 or a functional derivative or homologue thereof, said method comprising the steps of:
 - (a) providing a cell expressing UL15 and VP5;
 - (b) permitting a sample containing a test compound to come into association with the UL15-, VP5-expressing cell; and
 - (c) identifying a test compound which inhibits the function of the UL15/VP5 complex.
 - 26. An antiviral agent identified by any of the methods of claim 23 25.





(12)

EUROPEAN PATENT APPLICATION

(88) Date of publication A3: 09.09.1998 Bulletin 1998/37

(51) Int Cl.6: G01N 33/50, G01N 33/569

- (43) Date of publication A2: 26.08.1998 Bulletin 1998/35
- (21) Application number: 98301123.0
- (22) Date of filing: 16.02.1998
- (84) Designated Contracting States:
 AT BE CH DE DK ES FI FR GB GR IE IT LI LU MC
 NL PŢ SE
 Designated Extension States:
 AL LT LV MK RO SI
- (30) Priority: 21.02.1997 US 38914 P
- (71) Applicant: SMITHKLINE BEECHAM CORPORATION
 Philadelphia Pennsylvania 19103 (US)

- (72) Inventor: Del Vecchio, Alfred King of Prussia, PA 19406 (US)
- (74) Representative: Crump, Julian Richard John
 fJ Cleveland,
 40-43 Chancery Lane
 London WC2A 1JQ (GB)
- (54) Use of HSV-1 UL-15 and VP5 in identifying anti-viral agents
- (57) The present invention provides a method for identifying compounds having antiviral activity against members of the herpes virus family. This method involves the identification of compounds which inhibit the

interaction between the gene products of the herpes simplex virus-1 UL15 gene and the UL19 gene (VP5). Also provided are compounds identified by the method of the invention.



EUROPEAN SEARCH REPORT

Application Number

EP 98 30 1123

Category	Citation of document with it	ndication, where appropr	iate.	Releva	ant o	CLASSIFICATION OF THE
Calegory	of relevant pass			to clai		APPLICATION (Int.Cl.6)
A X	WO 92 03051 A (GENT * claim 8 *	A INC) 5 March	1992	1,2, 23-25 26		01N33/50 01N33/569
A	M.D. DAVISON ET AL: genes encoding two and VP26 of herpes JOURNAL OF GENENER vol. 72, 1992, GB, pages 2709-2713, XP * table 1 *	capsid protein simplex virus AL VIROLOGY,	s (VP24	1,2, 23-25		
A X	EP 0 566 554 A (MON 1993 * claim 25 *	ISANTO CO) 20 O	ctober	1,2, 23-25 26		
A X	WO 95 02071 A (UNIV 1995 * claim 1 *	PITTSBURGH) 1	_	1,2, 23-25 26		
A	MURATORE O ET AL: assay for antiviral herpes simplex viru MICROBIOLOGICA (PAV 257-261. ISSN: 0391	drug research s type 2." IA) 19 (3). 19 -5352, XP00206	on 96.	1,2, 23-25	L	TECHNICAL FIELDS SEARCHED (Int.CI.6)
X	* the whole documen	t *		26		
A	SUDO K ET AL: "A santiviral compounds virus type 1 using L929 cells." TOHOKU JOURNAL OF E 176 (3). 1995. 163-XP002069560	against herpe the MTT method XPERIMENTAL MEI	s simplex with DICINE	1,2, 23-25		
X	* the whole documen	t *		24		
		-,	/			
	The present search report has	been drawn up for all cla	ims			
	Place of search	Date of completion	on of the search			Examiner
	THE HAGUE	26 June	1998	\	dells,	, A
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EUROPEAN SEARCH REPORT

Application Number EP 98 30 1123

	DOCUMENTS CONSID	ERED TO BE RELEVA	NT						
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					TECHNICAL F SEARCHED	FIELDS (Int.Cl.6)			
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